

# Nutrients in Wastewater

Regional Training “Decentralized  
Wastewater Management”

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# Phosphorus

- Phosphorus is essential to the growth of algae and other biological organisms.
- The organically bound phosphorus is an important constituent of wastewater and sludge.

# Nitrogen

- Because nitrogen is an essential building block in the **synthesis of protein**, nitrogen data will be required to **evaluate the treat ability of wastewater by biological processes**.
- Insufficient nitrogen can necessitate the addition of nitrogen to make the wastewater treatable.
- Where control of alga growth in the receiving water is necessary to protect beneficial uses, removal or reduction of nitrogen in wastewaters prior to discharge may be desirable.
- The **total nitrogen**, as a commonly used parameter, consists of many numerous compounds such as;  $\text{NH}_3$ ,  $\text{NH}_4\text{-N}$ ,  $\text{NO}_3\text{-N}$ ,  $\text{NO}_2\text{-N}$ , urea, organic-N (amines, amino acids, ...etc).

# Why Test for Ammonia & Phosphorus

Wastewater may contain high levels of ammonia and phosphorus. Releasing these nutrients to a receiving stream may lead to eutrophication.

eutrophication - a process by which pollution from such sources as sewage effluent or leachate from fertilized fields causes a lake, pond, or fen to become over rich in organic and mineral nutrients, so that algae and cyanobacteria grow rapidly and deplete the oxygen supply

# Why Test for Ammonia & Phosphorus

Many new and renewal NPDES permits contain monitoring requirements for ammonia, phosphorus, and total nitrogen.

# Ammonia Sources

- Municipal wastewater treatment plants
- Agricultural runoff (fertilizers & manure)
- Industrial discharges (paper mills, mines, food processing)

# Phosphorus Sources

- Laundering or other cleaning
- Treatment of boiler waters
- Fertilizers (runoff)
- Water treatment chemicals

# Ammonia Analysis

*Standard Methods 4500 NH<sub>3</sub>*

- Titrimetric Method
- Ammonia Selective Electrode Method\*
- Phenate Method



# Ammonia Analysis

## Sample Preservation and Storage

- If samples will be analyzed within 24 hours, refrigerate at 4 °C
- If samples will be analyzed after 24 of collection, acidify to  $\text{pH} < 2$  with sulfuric acid and store at 4 °C.
- Preserved samples can be stored at 4 °C for up to 28 days.

# Ammonia by ISE

## Equipment

- pH/ISE Meter
- Ammonia ion selective electrode
- Stir plate
- Volumetric flasks
- Erlenmeyer flasks
- pipets

## Chemicals

- Ammonia standard (1000 ppm)
- 10 N sodium hydroxide

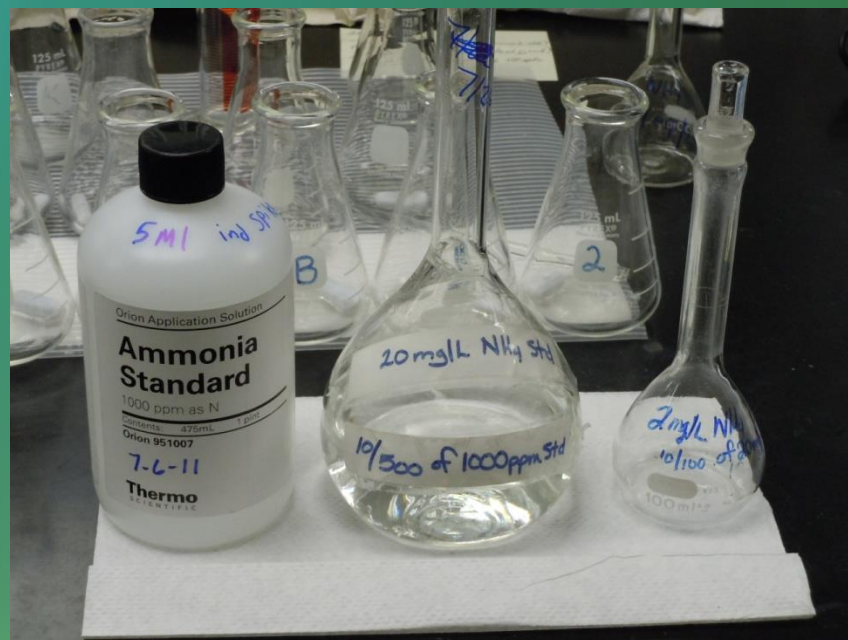
# Ammonia by ISE - Calibration

Prepare two standards.

The high standard should be 10 times the concentration of the low standard.

Use volumetric glassware.

*RRWRD uses 2.0 mg/L & 20.0 mg/L standards.*



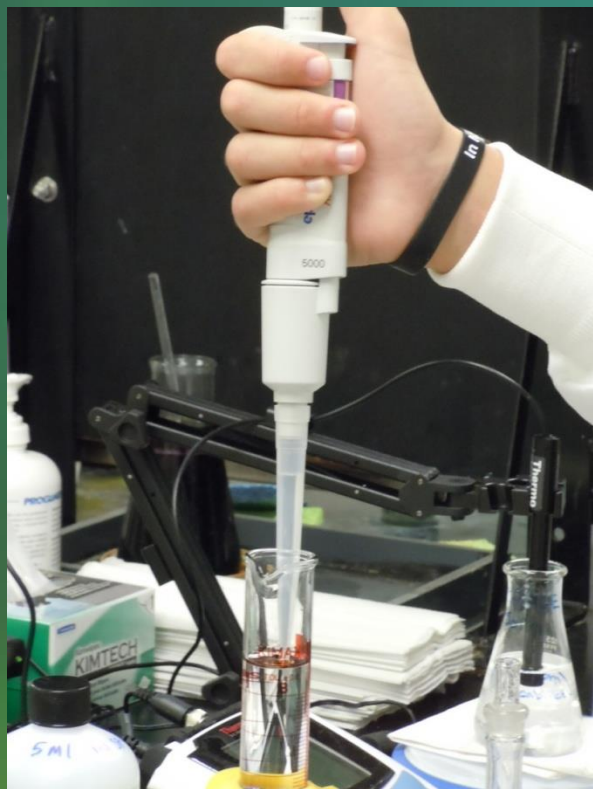
# Ammonia by ISE - Calibration

Follow directions supplied with your specific ion meter to calibrate meter.

## General Directions

- Pour 100 mL of each standard in an erlenmeyer flask.
- Place low standard flask on stir plate and stir moderately
- Insert electrode in sample
- Add 10 N NaOH.
- Allow meter to stabilize.
- Enter reading.
- Repeat for high standard





pipetting standard



diluting standard



transferring standard to  
flask





adding sodium hydroxide



waiting for meter to stabilize

# Ammonia by ISE - Calibration

Slope is the change in millivolts observed with every tenfold change in concentration.

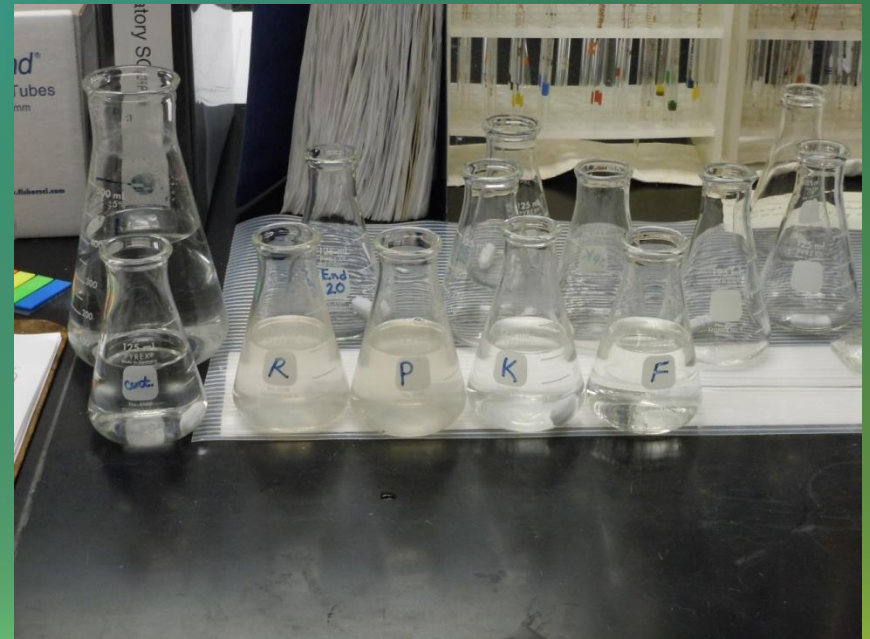
An acceptable slope is -54 to -60 at  $25 \pm 5^\circ\text{C}$ .

Do not analyze samples unless your slope is acceptable.

# Ammonia by ISE - Samples

## General Directions

- Pour 100 mL of sample into an Erlenmeyer flask.
- Place flask on stir plate and stir moderately
- Insert electrode in sample
- Add 10 N NaOH.
- Allow meter to stabilize.
- After meter has stabilized, record ammonia concentration.



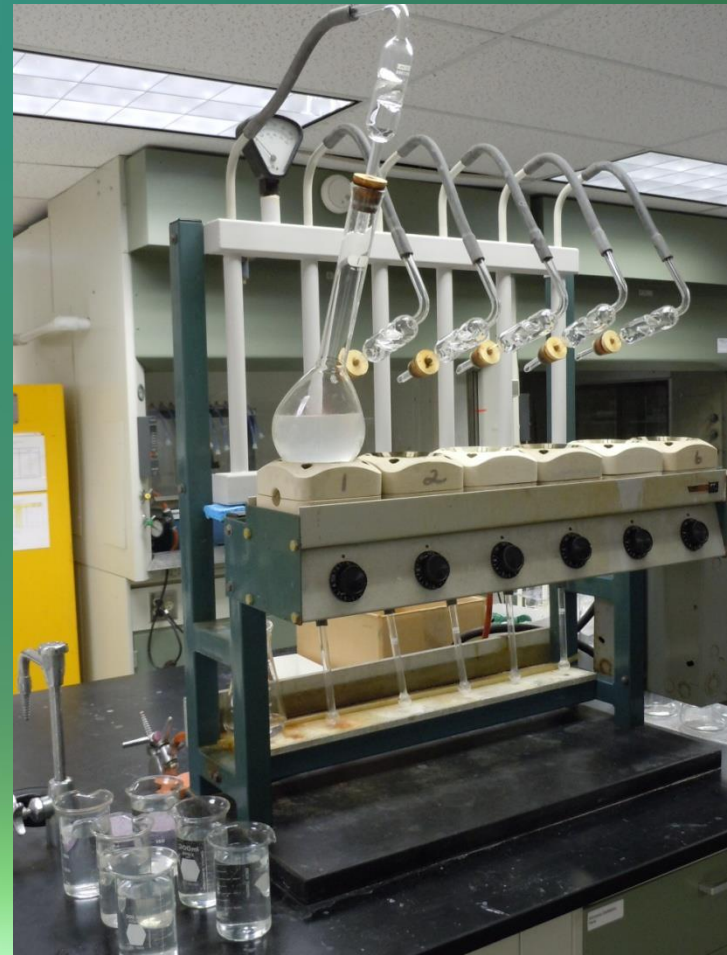


# Ammonia by ISE – Sample Pretreatment

You can eliminate interferences with preliminary distillation.

## General Directions

- Pour sample into Kjeldahl flask. Dilute if necessary
- Add borate buffer to sample
- Adjust pH to  $> 9.5$  with 6 N NaOH
- Attached flask to distillation apparatus.
- Distill.
- Collect distillate in 0.1% sulfuric acid.



# Ammonia by ISE – Sample Pretreatment



# Ammonia by ISE – solid samples

You can determine the ammonia present in sludge using pre-distillation.

Just weigh the sample before adding it to the Kjeldahl flask.

# Ammonia by ISE - Calculations

## Undistilled samples:

Meter reading \* 100 mL/sample volume

Distilled samples: assume sample volume = 500 mL

distillate volume = 100 mL

# Liquids

$$\text{Mtr reading} \times 500 \text{ mL} \times 100 \text{ mL}/(\text{mL spl} \times \text{mL distillate})$$

# Solids

# Ammonia by ISE – Analysis Range

## Low Level

- Use an approved EPA procedure to determine your method detection limit (MDL)  
(RRWRD MDL is 0.1)
- Your reporting limit is typically about 3 times your MDL.

## High level

- High end of range is your high standard.  
(RRWRD high is 20 mg/L.)
- You can extend high end of range by using a smaller sample volume and diluting it to 100 mL.

# Ammonia by ISE – Helpful Hints

- All samples and standards should be at the same temperature (preferably room temperature.)
- Use the same volume for standards and samples (usually 100 mL.)
- Use pH indicating ionic strength adjuster rather than 10 N NaOH
- Make sure readout is stable before recording ammonia concentrations. Samples with low ammonia levels may take more than five minutes to stabilize.
- Dirty samples can foul the electrode membrane.

# Phosphorus Analysis

Know what you want to analyze.

- Total reactive phosphorus
- Total acid–hydrolyzable phosphorus
- Total phosphorus
- Dissolved reactive phosphorus
- Dissolved acid-hydrolyzable phosphorus
- Dissolved phosphorus
- Suspended reactive phosphorus
- Suspended acid-hydrolyzable phosphorus
- Suspended phosphorus



# Phosphorus Analysis

## *Standard Methods 4500-P*

- Vanadomolybdophosphoric Acid Method (1 – 20 mg/L)
- Stannous Chloride Method (.01 – 6 mg/L)
- Ascorbic Acid Method (.01 – 6 mg/L)\*

*Hach 8190* – equivalent to *Standard Methods* 4500-P ascorbic acid method



# Phosphorus Analysis

If you want to analyze for any form of phosphorus other than total reactive phosphorus, you must pre-treat the sample.

Pretreatment may include

- Filtration
- Acid hydrolysis
- Digestion

# Total Phosphorus Analysis

## Sample Preservation and Storage

- Preservation – Adjust pH < 2 with hydrochloric acid
- Storage - Store at 4 °C for up to 28 days
- Note: Samples can leach phosphorus from plastic containers. Use glass containers for samples with low phosphorus levels.

# Total Phosphorus using Hach

(ascorbic acid colorimetry)

## Supplies

- Heating block (Hach COD reactor)
- Spectrophotometer
- Auto-pipetters

## Reagents

Hach Total Phosphorus Test N Tube Reagent Set includes reaction vials, potassium persulfate packets, sodium hydroxide, and ascorbic acid packets.

# Total Phosphorus – Hach Procedure

## General Directions

- Add 5 mL sample to digestion vial
- Add potassium persulfate powder. Shake to mix.
- Heat vial for 30 minutes
- Add 2 mL NaOH
- Add PhosVer 3 reagent
- Allow color to develop for 2 minutes
- Read concentration using spectrophotometer. Wavelength is 650.





adjusting auto-pipetter



pipetting samples



adding reagents

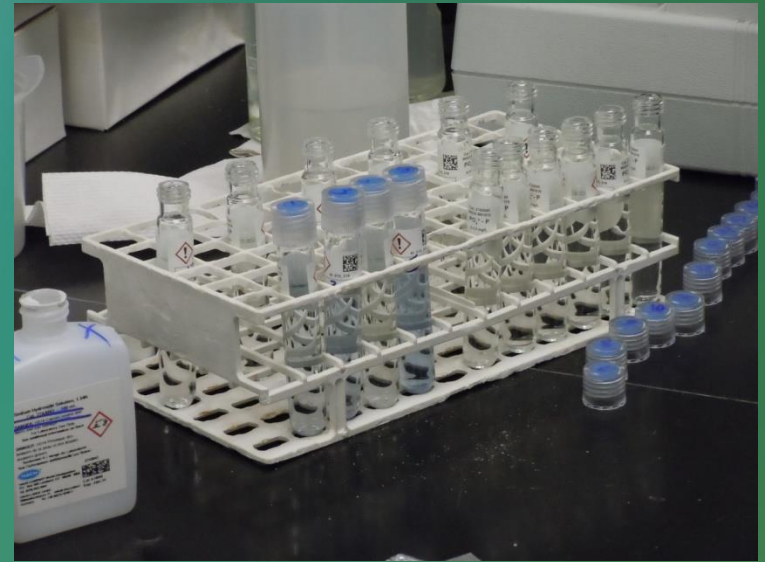


hot block digester





adding sodium hydroxide



shades of blue



spectrophotometer

# Total Phosphorus - Hach Procedure

## Calculations

Hach spectrophotometers will provide a readout in mg/L P.

Dilutions:

$$\text{mg/L P} * \frac{5 \text{ mL}}{\text{sample volume}}$$

# Total Phosphorus - Hach Procedure

## Analysis Range

Phosphorus: 0 – 1.10 mg/L P

Phosphate: 0 – 3.50 mg/L  $\text{PO}_4^{3-}$

You can extend the range by diluting samples before analysis.



# Total Phosphorus – Hach Procedure

## Helpful Hints

- Let heating block warm up for at least 10 minutes before inserting sample vials
- Shake samples well before pipetting sample aliquot.
- You can complete the procedure through the digestion step, and then finish it the next day.
- If there is sediment present in sample, make sure it is settled to bottom of tube before reading on the spectrophotometer.
- Read sample between 2 and 8 minutes of adding PhosVer3 powder pillow. This means you may have to be reading one sample while still adding reagent top others.

# Total Phosphorus for Solids

## (ascorbic acid colorimetry)

### Supplies

- Heating blocks
- UV/VIS Spectrophotometer
- Auto-pipetters
- Kjeldahl flasks
- Nessler's tubes

### Reagents

- Phenolphthalein Indicator
- Concentrated  $\text{H}_2\text{SO}_4$
- 10 N NaOH, Concentrated  $\text{HNO}_3$
- 5N  $\text{H}_2\text{SO}_4$
- Potassium Antimonyl Tartrate
- Ammonium Molybdate,
- Ascorbic Acid.



# Total Phosphorus – Solids

## Digestion

- Add sample to Kjeldahl flask and bring volume up to 200 mL with DI water. Add 2 mL of concentrated  $\text{H}_2\text{SO}_4$  and 5 mL concentrated  $\text{HNO}_3$ . Swirl to mix.
- Digest on a heating unit. Digestion is complete when solution is colorless and fumes are white. Cool.
- Add 20 mL DI water and few drops of phenolphthalein indicator. Add enough 10 N NaOH to produce a pink color.
- Quantitatively transfer solution to a 100 mL Volumetric flask.

# Total Phosphorus – Solids

## Colorimetry

- Pipet appropriate volume of digested samples into Nessler tubes. Add 5 N drop wise to discharge pink color. Adjust volume 50 mL with DI water.
- Add 8 mL of the color reagent and mix.
- Allow color to develop for 10 minutes but no longer than 30 minutes.
- Read concentration using spectrophotometer. Wavelength is 650.

$$\text{mg/Kg P} = \frac{\text{ug P}}{\text{mL colorimetry}} \times \frac{100 \text{ mL}}{\text{g spl} * \text{TS} * 0.1} \times \frac{1 \text{ mg}}{1000 \text{ ug}} \times \frac{1000 \text{ g}}{1 \text{ Kg}}$$

1.



2.



3.



1. weighing solid sample
2. adding water to sample
3. adding acid to sample

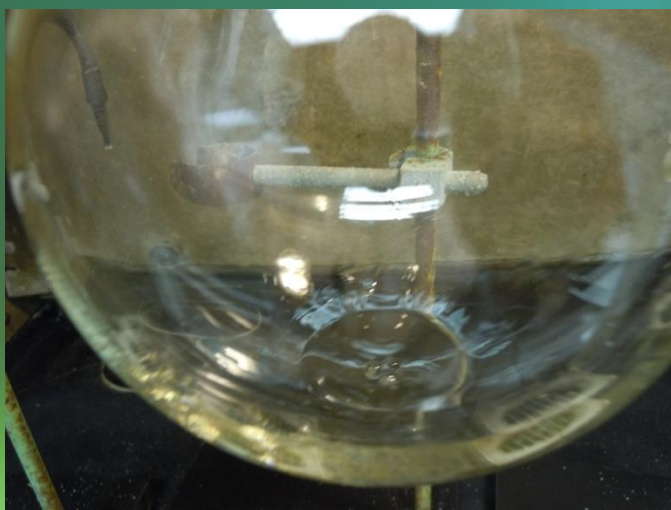




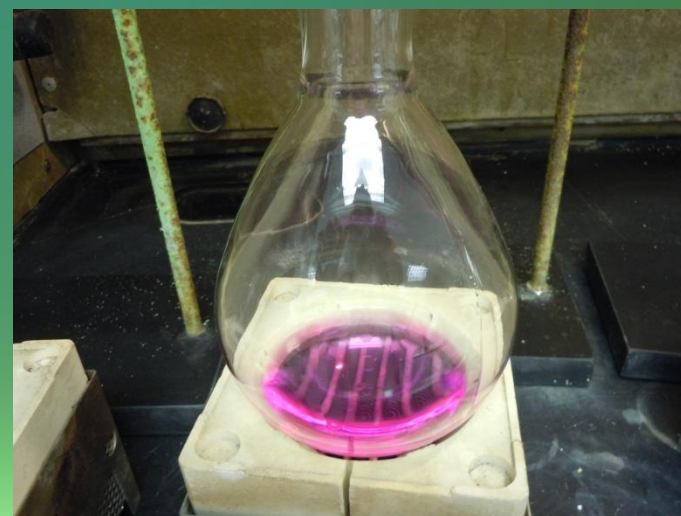
digesting samples



digestion almost complete



completely digested  
sample



pH adjusted digestate



1. Transferring sample from Kjeldahl to flask
2. Nessler tubes with pH adjusted samples
3. Samples after adding colorimetry reagents

# Quality Control

It is good practice to include the following in each analysis batch:

- Laboratory Control Sample (LCS) – this is a purchased solution of know concentration. Run it immediately after calibrating the meter.
- Blank – ammonia free water
- Matrix Spike Duplicate (MSD) – these are samples spiked with a know concentration of ammonia
- Continuing Calibration Verification (CCV) – this is usually a standard made in house. Run it after you read all the samples.



# Quality Control

How do you know if your analysis batch is “good?”

- LCS meets the manufacturer’s acceptance limits.
- Blank is less than half of the method detection limit.
- MSD recovery is 80 – 120%.
- CCV recovery is 90 – 110 %.

# Total Nitrogen

Many new and renewal permits include a Total Nitrogen monitoring requirement.

Total Nitrogen is organic nitrogen + ammonia + nitrate + nitrite

# Total Nitrogen Analysis

Typically labs determine total nitrogen by analyzing for Kjeldahl nitrogen (digestion followed by distillation) and nitrate/nitrite (using the cadmium reduction method.) These are traditional, wet lab analyses

Some labs use ion chromatography to analyze for total nitrogen.